## BIOMIMETIC SYNTHESIS AND STEREOSTRUCTURE OF K-13, A NOVEL INHIBITOR OF ANGIOTENSIN I CONVERTING ENZYME

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<u>Summary</u>: A novel inhibitor of angiotensin I converting enzyme (K-13) has been synthesized from N-acetyl-3,5-dichloro-L-tyrosyl-0-benzyl-L-tyrosyl-3,5-diiodo-L-tyrosine methyl ester, whose oxidation with thallium trinitrate (TTN) as a key step followed by zinc reduction affords the corresponding diphenyl ether with the same heterocyclic skelton as that of K-13, indicating that K-13 is biosynthesized from three molecules of L-tyrosine.

A novel inhibitor of angiotensin I converting enzyme, designated K-13, has been isolated from the culture broth of <u>Micromonospora halophytica</u> subsp. <u>exilisia</u> K-13,<sup>1</sup> and its structure has also been elucidated on the basis of the spectral data together with some chemical evidence.<sup>2</sup> However, the stereostructure of K-13 remains unsettled. In connection with OF 4949-III (1), an inhibitor of aminopeptidase B,<sup>3,4</sup> we are interested in the structure and physiological activity of K-13 (2), because their cyclization modes are different around a diphenyl ether moiety although their structures are similar to each other. We describe herein a total synthesis of K-13, indicating that its stereostructure is represented by 2.

3.5-Dibromo-L-tyrosine methyl ester was connected to N-t-butyloxycarbonyl-O-benzyl-Ltyrosine using DCC (1.2 equiv.) - 1-hydroxybenzotriazole (1 equiv.) - N-methylmorpholine (1 equiv.) in DMF (0 °C  $\rightarrow$  room temp., overnight) and then treated with HCl in dioxane (room temp., overnight) to afford the corresponding dipeptide (3),<sup>5</sup> in 70% overall yield. According to essentially the same procedure as described above, the desired tripeptide (4)<sup>5</sup> was further produced, in 78% yield, from 3 and N-t-butyloxycarbonyl-3.5-dichloro-L-tyrosine.

In the next step, the tripeptide (4) was oxidized with TTN (3 equiv.) in MeOH - dioxane



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(2:1) (0 °C  $\rightarrow$  room temp., overnight) and then treated with Zn in AcOH - THF (room temp., 3 h) to afford a cyclization product (5),<sup>6</sup> in 42% overall yield. Unfortunately, however, its cyclization mode is not identical with that of K-13 (2), but instead 5 has the same heterocyclic skelton as that of OF 4949-III (1). This macrocyclic compound (5) was subjected to catalytic hydrogenation [H<sub>2</sub>/Pd-black, NaOAc/MeOH (room temp., 2 days)] to give a phenol (6),<sup>6</sup> in 96% yield, which was then converted into the corresponding N-acetyl compound (7)<sup>6</sup> in 2 steps [1) HCl /dioxane (room temp., overnight); 2) Ac<sub>2</sub>O/pyr. (room temp., 5.5 h) (68% overall yield)]. Finally, 7 was hydrolyzed with 1N NaOH (3.1 equiv.) in MeOH (room temp., 25 min) and then treated with Amberlite IR-120 (H<sup>+</sup>) to afford an isomer (8) of K-13, in 98% yield. Its physiological property will be examined in due course.

As judged from the above result, 3,5-diiodo-L-tyrosine methyl ester must be used instead of the corresponding 3.5-dibromo compound. Thus, according to the same procedure as described above, 3,5-diiodo-L-tyrosine methyl ester was connected to N-t-butyloxycarbonyl-O-benzyl-L-tyrosine to give rise to the corresponding dipeptide (9),<sup>5</sup> in 97% overall yield, which was further reacted with N-acetyl-3,5-dichloro-L-tyrosine using DCC (1.2 equiv.) - 1-hydroxybenzo -triazole (1 equiv.) - N-methylmorpholine (1 equiv.) in DMF (0 °C  $\rightarrow$  room temp., 18 h) to afford a tripeptide (10)<sup>5</sup> in guantitative yield.

This compound (10) was subjected to TTN oxidation in MeOH – THF (1:2.5) (-22  $^{\circ}C \rightarrow$  room temp.. 9 h) followed by Zn in AcOH (room temp., overnight) to give a desired diphenyl ether  $(11)^6$  in 15% overall yield. In this case, another macrocyclic compound  $(12)^7$  was also obtained in ca. 5% yield, which was readily converted into 7 on catalytic hydrogenation. Finally, the macrocyclic diphenyl ether (11) was subjected to catalytic hydrogenation  $[H_2/Pd$ black, NaOAc/THF - MeOH (1:1) (room temp., 4 days)] to afford a desired phenol (13), in 20% yield. In this case, reductive dechlorination did not take place smoothly.<sup>8</sup> Furthermore, 13 was treated with 1N aq. NaOH (14 equiv.) and then with Amberlite IR-120 (H<sup>+</sup>) to give rise to the seventeen-membered macrocyclic tripeptide in quantitative yield. The synthetic tripeptide was completely identical with natural K-13 (1) in all respects of spectral data (IR and  $^{1}$ H NMR). Particularly, the optical rotation of the synthetic sample  $([\alpha]_0^{26}$  -7.4° (c 0.65, MeOH)) is quite similar to that of natural K-13 ( $\lceil \alpha \rceil_{D}^{19}$  -3.4° (c 0.62, MeOH)). According to essentially the same procedure as described above, the remaining three stereoisomers have also been synthesized.4.9 In connection with these tyrosine-derived antibiotics, further synthetic study on vancomycin and related compounds is in progress using our synthetic method demonstrated in the present paper.

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- 5. All new compounds described herein gave satisfactory spectral data consistent with the assigned structures.
- 6. The spectral data for the new compounds are in accord with the structures assigned, and only selected data are cited: 5: mp 151 154 °C;  $[a]p^{27} + 35.0^{\circ}$  (c 1.0, MeOH); IR (film) 3325, 2950, 1740, 1650, 1540, and 1500 cm<sup>-1</sup>;  $\delta$  (CDC1<sub>3</sub>) 1.50 (9H, s), 2.37 (1H, t, J= 12.7 Hz), 2.67 (1H, br.d., J= 14.2 Hz), 2.80 (1H, dd, J= 9.8, 16.1 Hz), 2.98 (1H, dd, J= 4.4, 14.2 Hz), 3.02 (1H, dd, J= 5.4, 14.2 Hz), 3.32 (1H, dd, J= 3.9, 13.4 Hz), 3.83 (3H, s), 4.41 (1H, m), 4.50 (1H, m), 4.94 (1H, m), 5.05 (2H, s), 5.41 (1H, d, J= 6.8 Hz), 5.67 (1H, d, J= 2.0 Hz), 5.91 (1H, s), 6.01 (1H, d, J= 9.8 Hz), 6.40 (1H, d, J= 7.8 Hz), 6.74 (1H, d, J= 2.0 Hz), 6.93 (2H, d, J= 8.8 Hz), 7.06 (1H, d, J= 2.0 Hz), 7.14 (2H, d, J= 8.8 Hz), 7.32 7.44 (5H, complex), and 7.60 (1H, d, J= 2.0 Hz). 6 as amorphus powder: C<sub>31H37N309</sub> [m/z 620 (M<sup>+</sup>+1)];  $[a]p^{28} + 43.0^{\circ}$  (c 1.25, MeOH); IR (Nujol) 3300, 1730, 1650, and 1500 cm<sup>-1</sup>;  $\delta$  (CD<sub>3</sub>OD) 1.43 (9H, s), 2.64 2.76 (3H, complex), 2.93 (2H, br.dd, J= 5.9, 13.9 Hz), 3.81 (3H, s), 4.23 (1H, complex), 4.53 4.56 (1H, complex), 4.78 (1H, m), 5.34 (1H, d. J= 8.3 Hz), 6.73 (1H, dJ= 2.0, 8.1 Hz), 6.63 (2H, d. J= 8.3 Hz), 6.73 (1H, d, J= 7.8 Hz), 6.34 (1H, dJ, J= 2.0, 8.1 Hz), 7.96 (1H, dJ, J= 6.63 (2H, d. J= 8.3 Hz), 6.73 (1H, dJ= 9.8 Hz), 7.36 (1H, dJ, J= 2.0, 8.1 Hz), 7.96 (1H, dJ, J= 6.99 7.01 (3H, complex), 7.18 (1H, dJ, J= 2.0, 8.1 Hz), 6.63 (2H, dJ= 10.5, 17.8 Hz), 6.63 (2H, dJ= 10.5, 17.8 Hz), 6.63 (2H, dJ= 10.5, 17.8 Hz), 6.84 (1H, dJ, J= 2.0, 8.1 Hz), 7.96 (1H, dJ, J= 6.99 7.01 (3H, complex), 7.18 (1H, dJ, J= 9.8 Hz), 7.36 (1H, dJ, J= 2.0, 8.1 Hz), 7.96 (1H, dJ, J= 6.99 7.01 (3H, complex), 7.18 (1H, dJ, J= 2.0 Hz), 5.8 (1H, dJ, J= 2.4, 18.7), 7.96 (1H, dJ, J= 2.4, 18.7), 7.90 (1H, dJ, J= 3.4, 13.2 Hz), 4.52 4.59 (2H, complex), 2.94 3.00 (2H, complex), 3.42 (1H, dJ, J= 3.4, 13.2 Hz), 4.52 4.59 (2H, complex), 4.77 (1H, dJ, J= 3.4, 13.2 Hz), 4.59 (1H, dJ, J= 7.7 Hz), 6.44 (1H, dJ, J= 1.5, 8.1 Hz), 6.62 (2H, d, J= 1.2 Hz), 6.80 (1H, dJ= 2.0, 8.3 Hz), 7.00 (1H, d
- 7. This compound has not been obtained in completely pure state, and directly used for the next experiment leading to the formation of 7.
- 8. Other methods for reductive dechlorination are examined.
- 9. These results will be published elsewhere in detail.

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